

# Development of sol-gel based carbon ceramic electrode modified by graphene oxide - polypyrrole nanocomposite for simultaneous determination of uric acid and dopamine in presence of ascorbic acid

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## Abstract

In this paper a sensitive carbon ceramic electrode (CCE) based on Methyltrimethoxy Silane (MTMOS) was prepared simply by three step sol-gel process. The surface of CCE was modified with polypyrrole -electrochemically reduced graphene oxide (ERGO-PPy) nanocomposite by electro polymerization. Scanning electron microscopy, electrochemical impedance spectroscopy, XRD, FT-IR and electrochemical techniques were used to characterize the composite electrode. The cyclic voltammetry and differential pulse voltammetry results suggested that the modified electrode exhibited enhanced sensitivity and selectivity towards the oxidation of dopamine and uric acid. Differential pulse voltammetry results show that simultaneous determination of DA and UA in presence of ascorbic acid could be performed selectively and sensitively at ERGO/PPy modified sol-gel based CCE. The oxidation peak currents of DA and UA in presence of AA, presented a good linear relationship with the concentrations in the range from 0.05 to 65  $\mu\text{M}$  and 0.1 to 120 $\mu\text{M}$  respectively with a detection limit of 0.002  $\mu\text{M}$  for DA and 0.08  $\mu\text{M}$  for UA. The good performance of the modified electrode provided a promising modified electrode in electrochemical determination of DA in human blood serum and urine samples.

**Keywords:** Sol-gel, Graphene oxide, Polypyrrole, dopamine, uric acid.

## 1. Introduction

The Sol-gel technology is a three-step process which includes the low temperature hydrolysis of a monomeric precursor of organometallic alkoxide and its condensation followed by poly-condensation to yield a polymeric oxo-bridged network [1]. The mid-1800s was the beginning of interest in sol-gel process with Ebelman and Graham's studies on silica gels [2]. For a period of time (last 1800 to 1920) sol-gel technology was one of the major field of activity among the researchers [3]. According to Dislich, at the beginning of the 80s of the 20<sup>th</sup> century, the number of publication shows the exponential growth of this claim [4]. In the permanent demand for ceramic and glasses with better properties and greater purity, three method of processing have proposed to have important roles. These are preparation of ultrafine powder of controllable size, ceramic from metal-organic polymers and the sol-gel process [3]. Low preparation temperatures and inorganic supports offer some advantages to sol-gel process over other methods, among which are chemical inertness, physical strength, and good surface renewability [5]. Additional advantages come from the fact that sol-gel derived materials are porous and thus mass transport is relatively easy through pores [6].

This advantageous cause sol-gel process to be one of the most used synthetic method in material science and nanotechnology [3] as the protective coatings, thin films, Nanoscale powders and Opto-mechanical materials [7]. Carbon-ceramic composite electrodes (CCEs) are comprised of a dispersion of carbon powder that is held together by sol-gel derived ceramic binder [8]. This composite material can be used in a wide range of application such as electrode in rechargeable batteries [9], supercapacitors [10] solar cells [11], fuel cells [12], Electrochemical sensors [5,6,8,13-18] and opto-plasmonic sensory devices. Since the CCEs in the first stage of preparation is a paste like material, so it is easy to shape and form it in any arbitrary structures and be used in a wide range of applications. However bare carbon ceramic electrodes as sensor have some drawbacks such as low detection sensitivity and poor resolution to specific analytes, therefore their modification is needed. Bulk modification of carbon-ceramic electrodes is an effective route which has been utilized by numerous authors [8,13-16], but this procedure is time consuming, expensive and difficult, where surface modification is not. However, some modifiers suffer from the weak adhesion and low physical stability on surface of this substrates but in presence of an immobilizing material, this procedure is the best way of modification [5]. In recent years, electrically conductive polymer (ECP) attracted great interest because of its hybrid properties of having metal-liked conductivity and polymeric characteristics [25] such as flexibility, low density, and ease of structural modification at the same times [26], leading to an enormous range of potential applications [27]. Among the ECPs, polypyrrole is popular due to its strong electrical properties, ease of synthesise and good environmental and mechanical stability [28], which rise to its

wide application in electronic devices, electrodes for rechargeable batteries [29], solid electrolytes for capacitors [30], sensors [31] and corrosion protection materials [28].

On the other hand, graphene, the building block of carbon allotropes, was theoretically established in 1940 [19]. Boehm and coworkers separated thin lamellae of carbon by heating and chemical reduction of graphite oxide in 1962 [20]. This discovery was a revolution in the fields of physics, chemistry, biotechnology, and materials science [21]. The “thinnest” known material graphene exhibits excellent electrical conductivity, mechanical flexibility optical transparency, thermal conductivity, and low coefficient of thermal expansion (CTE) behavior [22]. But most of this properties are only associated with individual sheets [23]. Among the graphene synthesis methods including mechanical and chemical exfoliations [23], epitaxial growth via chemical vapor deposition (CVD) [19] and unzipping of carbon nanotubes (CNT) [24], the oxidation of graphite by modified Hummers method, followed by chemical and electrochemical reduction, is the common method due to its simplicity and scalability [21,23].

Up to our knowledge, DA, UA and AA are always coexisted in the extra cellular fluids of central nervous system and serum in mammals [32], which play very important roles in human metabolism system [6]. DA, as one of the important neurotransmitters, belongs to the catecholamine group which belongs to the family of excitatory chemical neurotransmitters [5] and has very important role in the central nervous, renal, hormonal and cardiovascular systems [33]. Low level of DA may cause neurological disorders such as schizophrenia and Parkinson's disease [34]. Meanwhile, UA is the primary final product of purine metabolism [35]. Abnormal concentration of UA has a relation with several diseases such as hyperuricemia, gout and pneumonia [36]. AA is a common anti-oxidant, which plays a great role in the metabolic process of human bodies [36]. As the AA concentration is much higher (100–1000 times) than that of DA in body fluids, therefore selective detection of DA and UA, and even their simultaneous detection in the presence of large amount of AA have received tremendous attention in biomedical chemistry and diagnostic and pathological research owing to their similar properties [37]. As these molecules are electroactive, electrochemical methods are promising advantageous such as rapid response, high sensitivity, easy operation, and low cost [38]. However, the key challenge in determination of DA and UA, is the interference of coexisted AA with high concentrations, because AA can also be oxidized and its peak currents are overlapped with those of DA and UA at conventional electrodes [31]. Thus, a variety of modified electrodes were recently realized to either reduce the over-potential of AA oxidation or prevent the approach of AA to the electrode surface [32]. To deal with this problem, there is lots of modifiers materials which can facilitates determination of DA and UA even simultaneously by eliminating of AA reaction onto the electrode surfaces [32].

In this research (which is a part of the other project in which the presented electrochemical sensors will be coupled to a plasmonic and solitonic sensor), a CCE was fabricated by sol-gel process and graphene nano-sheets was immobilized on surface of CCE by electro-polymerization of pyrrole to enhance its electrochemical activity toward oxidation of DA and UA. This modified electrode was successfully used for simultaneous determination of DA and UA in the presence of high levels of AA in human blood serum and urine samples.

## **2. Experimental methods**

### **2.1. Apparatus**

Electrochemical experiment was carried out by AUTOLAB PGSTAT 30 electrochemical analysis system and GPES 4.9 software package (Eco Chemie, The Netherlands) consisting of three electrodes: bare or modified carbon ceramic electrode (CCE) as working electrode, Ag/AgCl as auxiliary electrode and a Pt wire as counter electrode. The FTIR spectroscopy was conducted on FT-IR Tensor 27 Bruker, over a frequency range of 4000- 500  $\text{cm}^{-1}$ . Crystallographic structure of graphene oxide and pure graphite were performed using a D/Max 2500V/PC diffractometer (Siemens-Germany). The scanning electron microscopy (SEM) experiment was made on a Hitachi Japan S4160 SEM.

### **2.2. Reagents and chemicals**

Methyltrimethoxy Silane (MTMOS), DA and graphite (GR) powder were purchased from Merck and were of analytical grade from Merck. Pyrrole monomer was purchased from Merck and distilled prior to use. Dopamine, ascorbic acid, uric acid and other reagents were of analytical grade from Merck. The human blood serum and urine samples were obtained from Drug Applied Research Center, Tabriz University of Medical Sciences-Iran. The pH of solutions was adjusted to 7.0 with phosphate buffer. Distilled, deionized and sterilized water was used in all solution preparations.

### 2.3. Preparation of graphene oxide

Modified Hummers method was used in preparation of graphene oxide [23]. Mixture of sulfuric acid (46 mL, 98 wt. %) and phosphoric acid (6 mL 85 wt. % in H<sub>2</sub>O) was poured to a mixture of graphite powder (2.0 g) and sodium nitrate (1.0 g) and cooled to 5 °C in ice bath. Potassium permanganate (6.0 g) was slowly added to prevent the temperature exceeding from 20 °C. The ice bath was removed and temperature was increased to 35±4 °C and kept for 12 hr. Deionized water (92 mL) was gradually added to the vessel and the temperature of vessel increased to 98 °C for 30 min. The heat was removed and reaction cooled by water bath for 10 min. The resultant brown-yellow suspension was diluted by additional of deionized water (280 mL) and treated with H<sub>2</sub>O<sub>2</sub> (20%, 50 mL) to remove the remained oxidizing agents. The suspension was washed repeatedly with 5% HCl and deionized water respectively. The resultant solid on the filter was dried at 60 °C (48 hr.) obtaining 2.7 g of GO.

### 2.4. Preparation of modified Carbon Ceramic Electrode

The carbon-ceramic electrode prepared as follows. The silica sol solution was prepared by mixing 0.6 mL of MTMOS, 0.9 mL methanol and 0.1 mL hydrochloric acid (0.1 M) and stirred for 5 min to afford a homogeneous gel solution. This ormosil was mixed well with 300 mg graphite powder for 5 min. The mixture was added to a Teflon tube (with 3 mm id and 3 cm length, and the length of composite in the tube was about 8 mm) and dried for 48 h at room temperature. The surface of all electrodes was removed by mechanical polishing with 800, 2000, 2500 and 3000 grit polishing papers, respectively. The electrodes were rinsed thoroughly with water to yield shiny surfaces. Copper wire contacted to the other end, provided the electrical contact. To modifying the surface of CCE, a mixture solution containing 1 mg/ml graphene oxide and 0.1M pyrrole monomer in phosphate buffer solution (pH=7) was prepared. Cyclic voltammetry (CV) methods was chosen to electro-polymerization of pyrrole monomer and subsequently electrodeposition of graphene oxide nano-sheets by sweeping the potential from 0 to 0.8 V for 5 scan (scan rate of 0.01 V/s). To reduction of graphene oxide in composite, CV method was used as an efficient tool. The reduction was performed in 0.1 M KCl solution and by double scanning of the potential from 0 to -1.2 V (scan rate of 0.05 V/s).

## 3. Results and discussion

### 3.1. Surface morphology of modified electrode

Scanning electron microscopy was used to characterize the surface of modified and bare electrodes. As shown in Fig. 1(c, d) on bare carbon ceramic electrodes a dense, scaly and porous surface but in the case of modified nanocomposite electrode a crispy, wrinkled and layer liked structure was observed. According to the Fig.1 (a,b), a thin layer of PPy was electrodeposited onto the surface of GO, that immobilizes the sheets on electrodes surface by hydrogen bonding between oxygenated functionality of GO and N-H of pyrrole ring that increases physical stability of modified electrode.

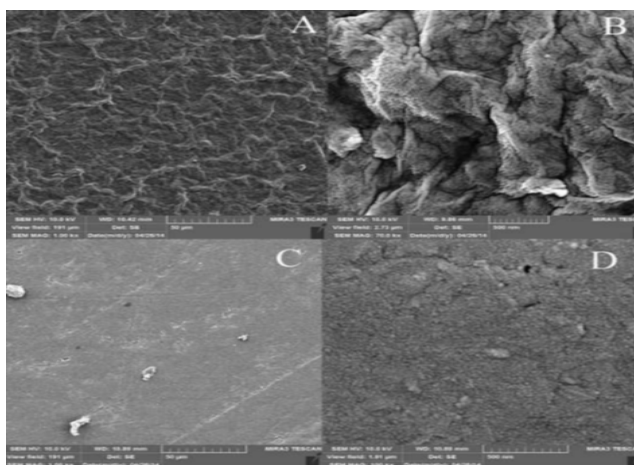


Figure 1. SEM images of (a, b) ERGO/CCE, (c, d) bare electrode with two different magnifications.

### 3.2. Characteristics of the modified electrodes by electrochemical impedance spectroscopy (EIS)

Electrochemical impedance spectroscopy can provide information about the impedance changes of the electrode surface by using  $[\text{Fe}(\text{CN})_6]^{3-/4-}$  redox couples as the electrochemical probe during the modification process. The Nyquist plots of modified and bare electrodes are shown in Fig. 2. The terms  $R_s$ ,  $\text{CPE}_1$ ,  $R_{ct}$  and  $W_1$  in the inset of Fig.2 refer to solution resistance, double layer capacity, charge transfer resistance and specific electrochemical element of diffusion or Warburg element, respectively. On the bare carbon-ceramic electrode, the charge transfers resistance ( $R_{ct}$ ), which was derived from the semicircle domains of impedance spectra, can be estimated to be  $1227.3 \Omega$  (curve a). On the ERGO/CCE (by casting of  $20 \mu\text{l}$ , GO with concentration of  $1 \text{ mg/ml}$  and electrochemical reduction) this value decreased to  $147 \Omega$  (curve b) which was attributed to high conductivity of ERGO on the surface of electrode. After electro-polymerization of polypyrrole and deposition of nanocomposite on the surface of carbon ceramic electrode, the  $R_{ct}$  value was increased to  $234 \Omega$  (curve c), that proves immobilization of nanocomposite on electrode surface. This decrease in conductivity was acceptable because, by introducing the PPy, the mechanical stability of ERGO on surface of carbon ceramic electrode was dramatically increased.

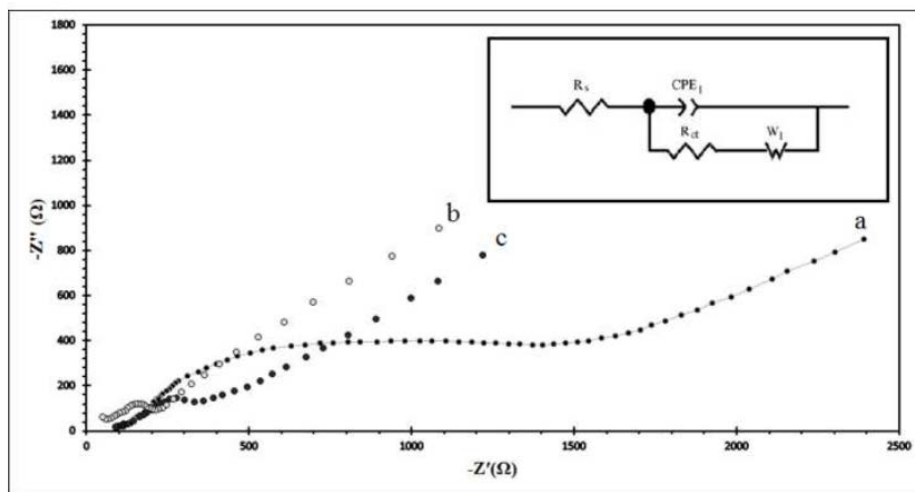


Figure 2. EIS for (a) bare carbon-ceramic electrode, (b) ERGO/ carbon ceramic electrode and (c) ERGOPPy/carbon ceramic electrode in a mixture solution of  $10.0 \text{ mM}$   $[\text{Fe}(\text{CN})_6]^{3-/4-}$  and  $0.1 \text{ M}$   $\text{KCl}$  with the frequencies swept from  $2 \times 10^6$  to  $0.1 \text{ Hz}$  under an open circuit potential condition.

### 3.3. XRD study

Although XRD is not a perfect tool for identifying single-layer graphene, but it is very informative. This analysis was used to verify the formation of the synthesized GO. According to Fig.3 the XRD pattern of pristine graphite shows that, after oxidation by modified Hummers' method, the peak around  $26.38^\circ$  corresponding to 002 reflections peak [19], shifts to the lower angles ( $11.36^\circ$ ). After oxidation of graphite, the interlayer distance of the sheets, increases from  $0.377 \text{ nm}$  (GR) to  $0.772 \text{ nm}$  (GO), proving oxidation of graphite. It reveals that the space of the sheets in GR increases by introducing the water molecules and generation of highly oxygenated functional group between the interlayer galleries. Generation of oxygen functionality attenuate  $\pi$ - $\pi$  interaction of the sheets and helps to better exfoliation of graphite oxide into graphene oxide.

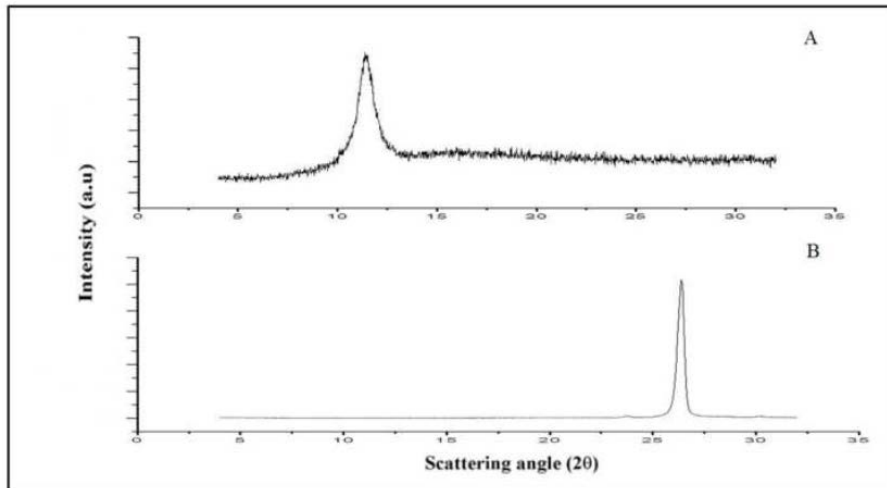


Figure 3. XRD structure of Graphite (A) and Graphene oxide synthesized by modified Hummers' method (B).

### 3.4. FT-IR spectroscopy

FT-IR spectroscopy is known to be an important tool to characterize the graphene based hybrid materials. From the Fig.4, the peaks at 3076 and 2882  $\text{cm}^{-1}$  are attributed to the asymmetric stretching and symmetric vibrations of  $\text{CH}_2$  [26]. The broad peak at 3432 and 1720  $\text{cm}^{-1}$  are designated to O-H stretching vibration and the carbonyl ( $\text{C}=\text{O}$ ) stretching respectively. Peak at 1059  $\text{cm}^{-1}$  proves the existence of epoxide group on GO sheets representing C-O stretching vibrations [39]. The peak appeared at 1620  $\text{cm}^{-1}$  is responsible for residual  $\text{C}=\text{C}$   $\text{sp}^2$  carbons and peak at 1179  $\text{cm}^{-1}$  attributed to C-C stretching vibrations [29].

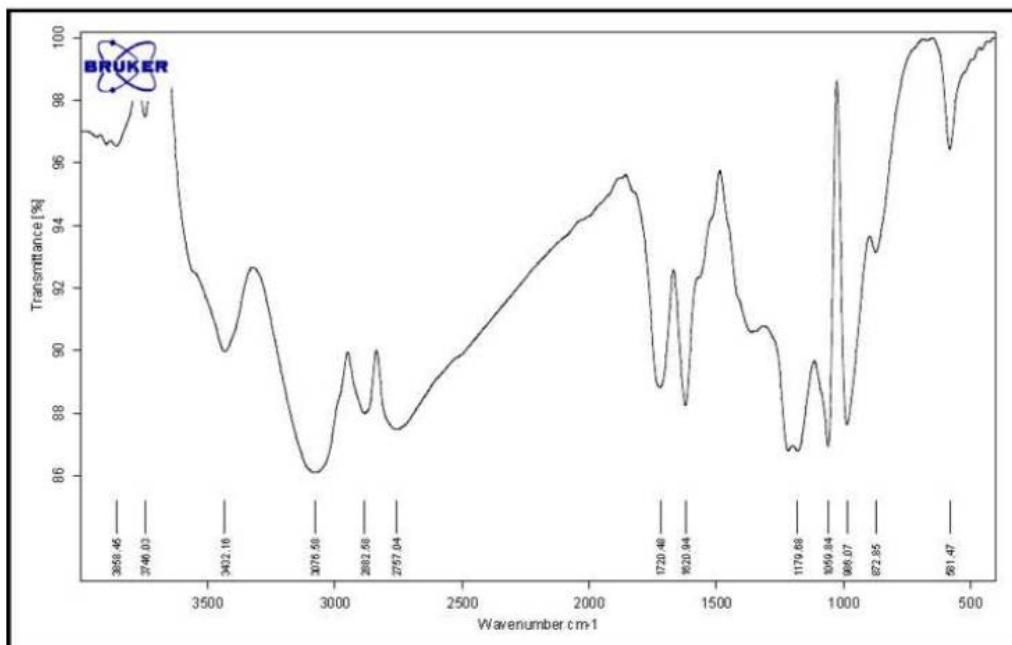


Figure 4. FT-IR analysis of Graphene oxide, synthesized by modified Hummers' method.



### 3.5. Cyclic voltammetry

Since Cyclic voltammetry (CV) has become an important and widely used electroanalytical technique in many areas of chemistry, it was selected for the assessing the ability of ERGO-PPy nanocomposite modified CCE toward catalytic oxidation of DA and UA in presence of AA. The cyclic voltammograms of bare and modified CCE are shown in Fig.5 (A). On the bare CCE DA, UA and AA shows broad anodic peak with the anodic peak potential of 229, 263 and 376 mV (Vs. Ag/AgCl) respectively. It is obvious that electron transfer kinetics in oxidation of these three species on bare CCE is sluggish and irreversible. In contrast modified CCE can effectively eliminate AA electrochemical reaction and separate overlapped oxidation peaks of DA and UA. For DA on bare CCE oxidation and reduction peaks appears at 239 and 150 mV ( $\Delta E=89$  mV), while in case of ERGO/PPy/CCE (Fig.5 (B)) it appears at 228 and 180 mV ( $\Delta E=48$  mV) respectively. Oxidation current of DA on ERGO/PPy/CCE is 3.88 times higher than that of bare CCE which suggesting electro-catalytic activity of ERGO/PPy toward DA oxidation. In case of UA on bare and modified CCE oxidation peaks appears at the same potential but oxidation currents of modified CCE toward UA is 6 times higher than that of unmodified which suggest ERGO/PPy can increase in access surface area of bare CCE and as a result, increase the sensitivity of electrode. Cyclic voltammograms of bare and modified CCE in a ternary system containing 1mM of DA UA and AA in phosphate buffer solution are shown in Fig.5 A and B (curves d). Results shows that on bare CCE there is no enough resolution to determine DA and UA in presence of AA, but in case if modified CCE by eliminating the AA responses, resolution of electrode improves and makes simultaneous determination of DA and UA possible.

To evaluate the reproducibility of the modified electrode in simultaneous detection of DA and UA, a ternary solution of 1 mM DA, 1mM UA and 1mM AA was detected through an operation of 50 cycles in a potential ranging from -0.2 to 0.9 V. The absence of any remarkable change of the voltammetric current response on modified electrode revealed that it has excellent electrochemical stability. The related standard deviation (RSD) of the oxidation peak currents of DA and UA are used to assess the reproducibility of the detection. The RSDs of the peak currents of 50 successive runs are 2.23% and 3.11% for DA and UA, respectively, which are both lower than 4%, showing excellent reproducibility of the ERGO/PPy/CCE. We believe the porous nature of the carbon ceramic electrode allows ERGO-PPy be adsorbed by CCE and this prevents its tubing from the surface of CCE.

The electrochemical behavior of DA and UA toward ERGO- PPy/CCE was investigated in phosphate buffer (0.1M) with different pH using cyclic voltammetry. The catalytic oxidation peak of DA and UA shifted to less positive potential by increasing the pH values with linear slope of 53 and 56 mV/pH, which proves the proton involvement in the overall mechanism of electron transfer. As the anodic current has maximum value at pH 7. This pH was chosen as the optimum value for later experiments.

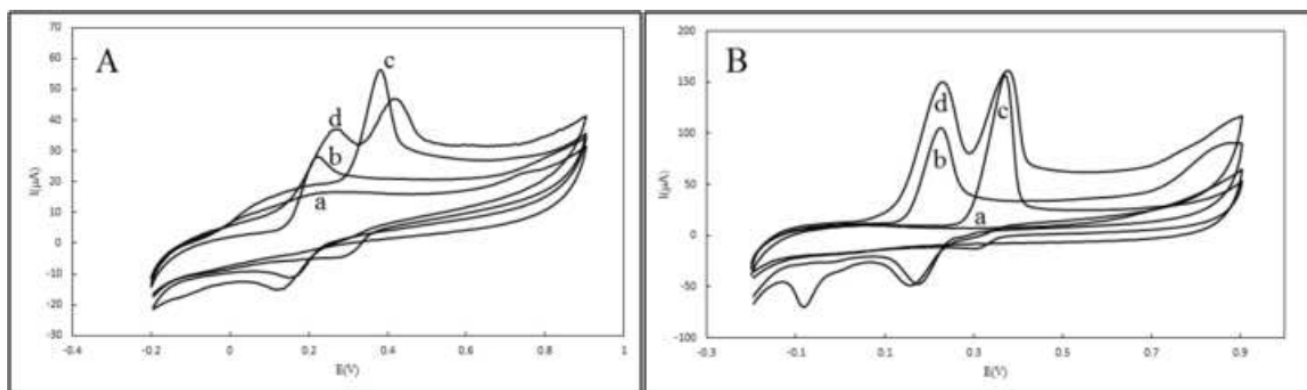


Figure 5. A: Cyclic voltammograms of bare CCE (a) in presence of 1mM AA, (b) 1mM AA and DA, (c) 1mM AA and 1mM UA and (d) 1mM AA, 1mM DA and 1mM UA. B: cyclic voltammograms of ERGO/PPy modified CCE in presence of (a) in presence of 1mM AA, (b) 1mM AA and DA, (c) 1mM AA and 1mM UA and (d) 1mM AA, 1mM DA and 1mM UA.

### 3.6. Differential pulse voltammetry study

Pulse voltammetric techniques, such as differential pulse voltammetry, have been recognized as useful tools in the rapid and sensitive detection in electroanalytical chemistry. Simultaneous detection of DA and UA was performed on the ERGO/PPy/CCE in 0.1 M phosphate solutions containing 1.0 mM AA. As shown in Fig. 6A, with the increase of the DA concentrations in the presence of 1.0 mM AA and 0.1 mM UA, the anodic peak currents of DA are linearly increased ranging from 0.05 to 65  $\mu\text{M}$ . For DA the regression equation is  $y = 0.4396C + 1.2109$ . Similar trends are observed by increasing the UA concentrations (Fig.6B) from 0.1 to 120.0  $\mu\text{M}$  (regression equation is  $y = 0.2921C + 0.9021$ ) in the solution containing 1.0 mM AA and 0.1 mM DA. The detection limits for DA and UA are evaluated to be about 0.002 and 0.08  $\mu\text{M}$  at 3 folds of the signal to noise ratio ( $S/N = 3$ ), respectively.

Fig.6C shows simultaneous determination of DA and UA in presence of 1mM AA. By increasing the concentration of DA and UA simultaneously, oxidation peak currents linearly increase accordingly. The linear range is from 0.4 to 80 $\mu\text{M}$  for DA ( $y = 0.8809C - 0.0986$ ) and from 0.4 to 50  $\mu\text{M}$  for UA ( $y = 0.9221x - 0.0787$ ), respectively. The detection limits are found to be 0.12  $\mu\text{M}$  for DA and 0.16  $\mu\text{M}$  for UA at three folds of the signal-to-noise ratio ( $S/N = 3$ ).

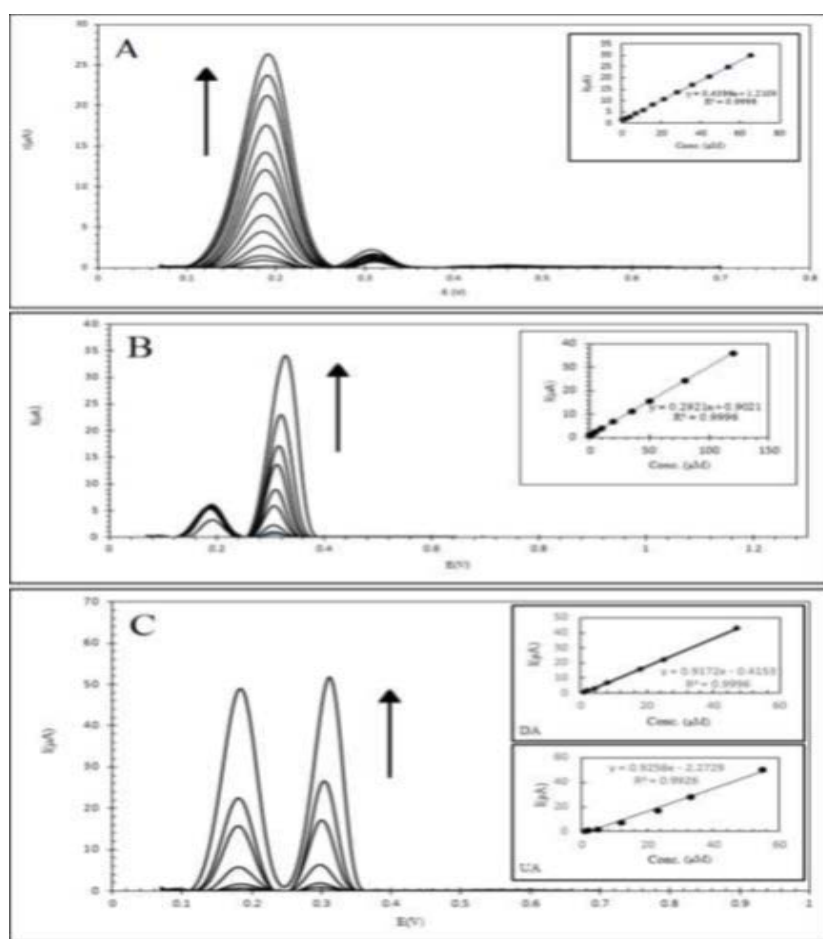


Figure 6. A: DPV of the ERGO/PPy/CCE in 0.1 M phosphate solutions (pH 7.0) containing 1.0 mM AA and 0.1 mM UA in the presence of different concentrations of DA: 0.1, 1.0, 1.9, 3.9, 6.8, 10.7, 15.6, 21.4, 28.1, 35.7, 44.2, 53.6 and 65.8  $\mu\text{M}$ . (B) DPV of the ERGO/PPy/CCE in 0.1 M phosphate solutions (pH 7.0) containing 1.0 mM AA and 0.1 mM DA in the presence of different concentrations of UA: 0.2, 1.0, 2.0, 5.0, 10.0, 20.0, 35.0, 50.0, 80.0 and 120  $\mu\text{M}$ . (C) DPV of the ERGO/PPy/CCE in 0.1 M phosphate solutions (pH 7.0) containing 1. mM AA in the presence of different concentrations of DA: 0.4, 2.0, 4.0, 8.0, 18.0, 25.0 and 47.0  $\mu\text{M}$  and UA 0.4, 2.0, 5.0, 12.0, 23.0, 33.0 and 55.0  $\mu\text{M}$ . Insets show the calibration curves of the DA and UA concentration vs. the anodic peak currents. Experimental conditions: step potential: 0.005 V, modulation amplitude: 0.025 V, scan rate: 0.01 V s<sup>-1</sup>.

Table 1 compares the response characteristics for simultaneous determination of DA and UA at various modified electrodes. In comparison with other modified electrodes [35-38,40], the proposed modified electrode possesses unique properties. Firstly, it has all the advantages of CCE and benefited from the advantages comes from mechanical properties of PPy and electrical properties of ERGO. The proposed modified electrode can be easily constructed in a simple procedure, and the surface of electrode is also easily renewable. Also the modified electrode shows high conductivity and promotion of fast electron transfer. Secondly, the ERGO-PPy/CCE eliminates the interference of AA in DA and UA determination due to proposed interaction mentioned above. Furthermore, sensitivity, detection limit and linear range of the proposed electrode are comparable with other methods.

*Table 1. Response properties of determination of DA at various modified electrodes*

Electrode	Method	Linear range (μM)		Detection limit (μM)	Refs.
		DA	UA		
layer-by-layer graphene and chitosan assembled multilayer/GCE	DPV	0.1 - 140.0	0.050	0.100	[37]
		1.0 - 125.0	0.100		
pretreated pencil graphite electrode/GCE	DPV	0.15 - 15.0	0.033	0.120	[38]
		0.30 - 150.0	0.120		
poly(L-methionine)/gold nanoparticle GCE	DPV	0.05 - 5.0	0.037	0.045	[35]
		0.07 - 7.0	0.045		
poly (naphthol green B)-film modified/CPE	DPV	5.0 - 270.0	0.250	5.0	[40]
		12.5 - 750.0	5.0		
Pt nanoparticles supported on reduced graphene oxide/GCE	DPV	10.0 - 170.0	0.250	0.450	[36]
		10.0 - 130.0	0.450		
Electrochemically reduced graphene oxide/polypyrrole/CCE	DPV	0.05 - 65.0	0.002	0.08	This work
		0.1 - 120.0	0.08		

### 3.7. Application

The ERGO-PPy modified carbon ceramic electrode was used for the determination of DA and UA in human blood serum and urine samples (Table 2). 1.0 mL human blood serum sample was deproteinized by adding 2.0 mL of acetonitrile. After centrifugation at 12000 rpm for 20 min, the supernatant was transferred into another centrifuge tube and dried with an N<sub>2</sub> stream. The residue was dissolved in 1.0 mL of water. Urine samples was diluted with 0.1M PBS (pH=7) and appropriate amount of these diluted samples were transferred to the electrochemical cell for the determination of each species by DPV method. The standard addition method was used for testing recovery. The analytical results summarized in table 2, which clearly indicate the potential application of ERGO/PPy/CCE for determination of DA and UA in real sample.

*Table 2. Experimental results for the determination of DA and UA in samples.*

Sample	Added (μM)	Found <sup>a</sup> (μM)	Recovery (%)
Human blood serum 1 <sup>b</sup>	-	-	-
	10	10.2 ± 0.3	102
Human blood serum 2 <sup>b</sup>	-	-	-
	20	20.6 ± 0.4	103
Urine 1 <sup>b</sup>	-	36.3 ± 0.9	-
	20	55.8 ± 0.5	97.5
Urine 2 <sup>b</sup>	-	63.2 ± 0.7	-
	40	102.4 ± 0.4	98

<sup>a</sup> Mean value ± standard deviation (n = 3).

<sup>b</sup> The dilution factor is 10 for human blood serum and 50 for urine sample.



#### 4. Conclusion

In this work we have fabricated a homemade CCE based on MTMOS modified with ERGO-PPy nanocomposite simply by electro-polymerization. The XRD analysis and FT-IR spectroscopy further confirmed oxidation of graphite and synthesizing of GO. The electrochemical reduction of GO composite was investigated in 0.1 M KCl at 0 to -1.2 V by 3 scan at scan rate of 100mV/s. This modified electrode was applied successfully as DA and UA sensing platform and the electrochemical experiments showed that this modified electrode can determine DA and UA simultaneously in presence of AA with high concentration level. Based on this method, a new biosensor was successfully developed for determination of DA and UA in human blood serum and urine samples.

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#### References

1. Sakka S (2005) Handbook of sol-gel science and technology. 1. Sol-gel processing, vol 1. Springer,
2. Hench LL, West JK (1990) The sol-gel process. Chemical Reviews 90 (1):33-72. doi:10.1021/cr00099a003
3. Mackenzie JD (1988) Applications of the sol-gel process. Journal of Non-Crystalline Solids 100 (1):162-168
4. Dimitriev Y, Ivanova Y, Iordanova R (2008) History of sol-gel science and technology. Journal of the University of Chemical technology and Metallurgy 43 (2):181-192
5. Majidi MR, Baj RFB, Naseri A (2013) Carbon nanotube-ionic liquid nanocomposite modified carbon ceramic electrode for determination of dopamine in real samples. Central European Journal of Chemistry 11 (7):1172-1186
6. Majidi MR, Asadpour H, Zeynali K, Ghobadipour S (2010) Nanobiocomposite of Polypyrrole/Carbon Nanotube/TiO<sub>2</sub>- Plant Tissue. Journal of Electroanalytical Chemistry 686 (1):177-182
7. Brinker CJ, Scherer GW (1990) Sol-gel science: the physics and chemistry of sol-gel processing. Gulf Professional Publishing,
8. Majidi MR, Asadpour H, Zeynali K, Ghobadipour S (2010) The Inhibitory Effect of Atrazine on Polyphenol Oxidase Activity for Determination of Atrazine. Journal of the Chinese Chemical Society 55 (3):522-528
9. Lee KT, Jung YS, Oh SM (2003) Synthesis of tin-encapsulated spherical hollow carbon for anode material in lithium secondary batteries. Journal of the American Chemical Society 125 (19):5652-5653
10. Portet C, Taberna P, Simon P, Laberty-Robert C (2004) Modification of Al current collector surface by sol-gel deposit for carbon-carbon supercapacitor applications. Electrochimica Acta 49 (6):905-912
11. Joshi P, Xie Y, Ropp M, Galipeau D, Bailey S, Qiao Q (2009) Dye-sensitized solar cells based on low cost nanoscale carbon/TiO<sub>2</sub> composite counter electrode. Energy & Environmental Science 2 (4):426-429
12. Anderson ML, Stroud RM, Rolison DR (2002) Enhancing the activity of fuel-cell reactions by designing three-dimensional nanostructured architectures: catalyst-modified carbon-silica composite aerogels. Nano Letters 2 (3):235-240
13. Salimi A, Alizadeh V, Hadadzadeh H (2004) Renewable Surface Sol-gel Derived Carbon Ceramic Electrode Modified with Copper Complex and Its Application as an Amperometric Sensor for Bromate Detection. Electroanalysis 16 (23):1984-1991
14. Salimi A, Pourbeyram S (2003) Renewable sol-gel carbon ceramic electrodes modified with a Ru-complex for the amperometric detection of l-cysteine and glutathione. Talanta 60 (1):205-214
15. Sheng Q, Yu H, Zheng J (2007) Sol-gel derived terbium hexacyanoferrate modified carbon ceramic electrode: Electrochemical behavior and its electrocatalytical oxidation of ascorbic acid. Journal of Electroanalytical Chemistry 606 (1):39-46
16. Wang P, Wang X, Zhu G (2000) Sol-gel-derived carbon ceramic composite electrodes bulk-modified with 1: 12-silicomolybdic acid. New journal of chemistry 24 (7):481-483

17. Caldas E, de Menezes E, Pizzolato T, Dias S, Costa T, Arenas L, Benvenuti E (2014) Ionic silsesquioxane film immobilized on silica applied in the development of carbon paste electrode for determination of methyl parathion. *J Sol-Gel Sci Technol* 72 (2):282-289. doi:10.1007/s10971-014-3367-8
18. Chen J, Zhang T, Gao C, Zhao Y (2010) Modifying glassy carbon electrode with ferrocene-bridged polysilsesquioxanes. *J Sol-Gel Sci Technol* 55 (3):293-298. doi:10.1007/s10971-010-2248-z
19. Kuila T, Bose S, Mishra AK, Khanra P, Kim NH, Lee JH (2012) Chemical functionalization of graphene and its applications. *Progress in Materials Science* 57 (7):1061-1105
20. Singh V, Joung D, Zhai L, Das S, Khondaker SI, Seal S (2011) Graphene based materials: past, present and future. *Progress in Materials Science* 56 (8):1178-1271
21. Dreyer DR, Park S, Bielawski CW, Ruoff RS (2010) The chemistry of graphene oxide. *Chemical Society Reviews* 39 (1):228-240
22. Brownson DA, Banks CE (2010) Graphene electrochemistry: an overview of potential applications. *Analyst* 135 (11):2768-2778
23. Park S, Ruoff RS (2009) Chemical methods for the production of graphenes. *Nature nanotechnology* 4 (4):217-224
24. Seger B, Kamat PV (2009) Electrocatalytically active graphene-platinum nanocomposites. Role of 2-D carbon support in PEM fuel cells. *The Journal of Physical Chemistry C* 113 (19):7990-7995
25. Layek RK, Nandi AK (2013) A review on synthesis and properties of polymer functionalized graphene. *Polymer* 54 (19):5087-5103
26. Bose S, Kuila T, Uddin ME, Kim NH, Lau AK, Lee JH (2010) In-situ synthesis and characterization of electrically conductive polypyrrole/graphene nanocomposites. *Polymer* 51 (25):5921-5928
27. Liu J, Wang Z, Xie X, Cheng H, Zhao Y, Qu L (2012) A rationally-designed synergetic polypyrrole/graphene bilayer actuator. *Journal of Materials Chemistry* 22 (9):4015-4020
28. Hosseini M, Sabouri M, Shahrabi T (2006) Comparison between polyaniline-phosphate and polypyrrole-phosphate composite coatings for mild steel corrosion protection. *Materials and Corrosion* 57 (5):407-410
29. Wang X, Yang C, Li H, Liu P (2013) Synthesis and electrochemical performance of well-defined flakes shaped sulfonated graphene/polypyrrole composites via facile in situ doping polymerization. *Electrochimica Acta* 111:729-737
30. Chang H-H, Chang C-K, Tsai Y-C, Liao C-S (2012) Electrochemically synthesized graphene/polypyrrole composites and their use in supercapacitor. *Carbon* 50 (6):2331-2336
31. Gao F, Cai X, Wang X, Gao C, Liu S, Gao F, Wang Q (2013) Highly sensitive and selective detection of dopamine in the presence of ascorbic acid at graphene oxide modified electrode. *Sensors and Actuators B: Chemical* 186:380-387
32. Han D, Han T, Shan C, Ivaska A, Niu L (2010) Simultaneous Determination of Ascorbic Acid, Dopamine and Uric Acid with Chitosan-Graphene Modified Electrode. *Electroanalysis* 22 (17-18):2001-2008
33. Majidi MR, Asadpour-Zeynali K, Gholizadeh S (2009) Amperometric biosensor for dopamine determination based on over-oxidized polypyrrole-plant tissue composite. *International Journal of Polymer Analysis and Characterization* 14 (2):89-101
34. Deng M, Yang X, Silke M, Qiu W, Xu M, Borghs G, Chen H (2011) Electrochemical deposition of polypyrrole/graphene oxide composite on microelectrodes towards tuning the electrochemical properties of neural probes. *Sensors and Actuators B: Chemical* 158 (1):176-184
35. Ojani R, Raof J-B, Maleki AA, Safshekan S (2014) Simultaneous and sensitive detection of dopamine and uric acid using a poly(L-methionine)/gold nanoparticle-modified glassy carbon electrode. *Chinese Journal of Catalysis* 35 (3):423-429.
36. Xu T-Q, Zhang Q-L, Zheng J-N, Lv Z-Y, Wei J, Wang A-J, Feng J-J (2014) Simultaneous determination of dopamine and uric acid in the presence of ascorbic acid using Pt nanoparticles supported on reduced graphene oxide. *Electrochimica Acta* 115 (0):109-115.
37. Weng X, Cao Q, Liang L, Chen J, You C, Ruan Y, Lin H, Wu L (2013) Simultaneous determination of dopamine and uric acid using layer-by-layer graphene and chitosan assembled multilayer films. *Talanta* 117 (0):359-365.
38. Alipour E, Majidi MR, Saadatirad A, Alizadeh AM (2013) Simultaneous determination of dopamine and uric acid in biological samples on the pretreated pencil graphite electrode. *Electrochimica Acta* 91:36-42
39. Wang P, Zheng Y, Li B (2013) Preparation and electrochemical properties of polypyrrole/graphite oxide composites with various feed ratios of pyrrole to graphite oxide. *Synthetic Metals* 166:33-39
40. Chittravathi S, Kumara Swamy BE, Mamatha GP, Sherigara BS (2012) Electrochemical behavior of poly

(naphthol green B)-film modified carbon paste electrode and its application for the determination of dopamine and uric acid. *Journal of Electroanalytical Chemistry* 667 (0):66-75

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